

Assessment of manila clam larval survival and physiological changes at 400, 520, and 1000ppm pCO₂ treatments David C. Metzger^{*1}, Shallin Busch², Paul McElhany², Carolyn S. Friedman¹, Steven B. Roberts¹

Introduction

Ocean Acidification as a result of increasing levels of dissolved CO₂ has been shown to impact the survival, physiology, and morphology of calcifying organisms.

Larval stages are thought to be at particular risk among bivalve species due to their dependences on soluble calcium concentrations

Limited studies exist that focus on the transcriptional response of calcifying organisms as a result of ocean acidification

Goals

1. Assess the impacts of elevated pCO_2 treatments on clam larval survival and morphology.

2. Identify changes in transcriptome as a result of elevated pCO_2 treatments.

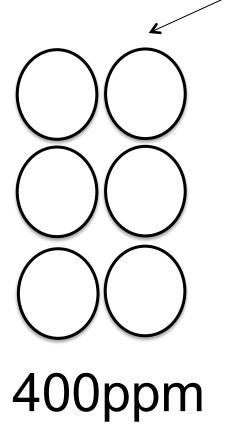
3. Characterize physiological changes at the molecular level as a result of elevated pCO2 conditions.

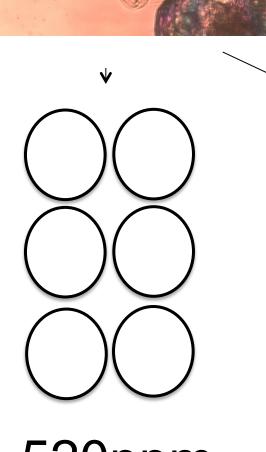
Experimental Design

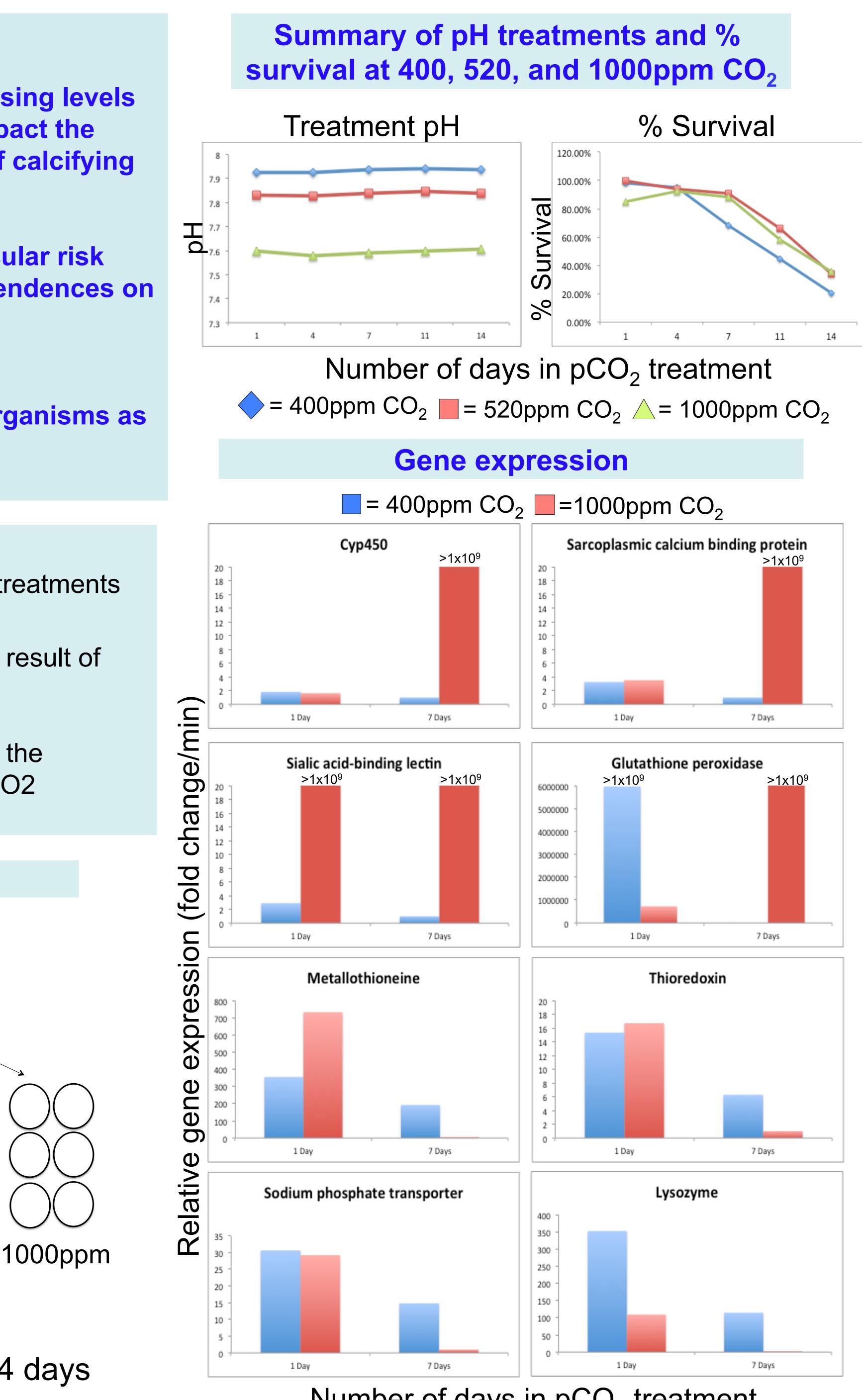
5 day old larvae

Split into three pCO2 treatments with 6 replicates/ treatment

Take samples for mortality, morphometrics, and qPCR







520ppm

1, 4, 7, 11, and 14 days

¹School of Aquatic and Fishery Sciences, University of Washington, Seattle, WA., ²Northwest Fisheries Science Center, NOAA, Seattle, WA

Number of days in pCO₂ treatment

Conclusions

Elevated pCO₂ levels appear to have no impact on 5 day old larval clam survival. Decreased survival at 400ppm (ambient) may have been a result of ciliate contamination in the system.

Gene expression varies widley depending on the phsyiological function.

Genes associated with stress response and ion transport are dramatically induced after one week of treatment at 1000ppm CO₂

Future Directions

Generate transcriptome libraries from 1 week samples at 400 and 1000ppm CO₂ treatments.

Further validation of qPCR results. More replicates and more timepoints.

Sequence and measure transcripts of genes identfied in other organisms (ie sea urchins) that are thought to be impacted by ocean acidification.

Complete assessment of larval growth rates under different pCO2 conditions.

Acknowledgements







Special thanks to Taylor Shellfish farms, NOAA's Northwest Fisheries Science Center, and the University of Washington School of Aquatic and Fisheries Sciences for assistance with this research. This work was funded by the Washington Sea Grant and NOAA's Saltonstall-Kennedy program.



